A Review on the Historical Development of Phytoplankton in the Philippines and their Biological Importance throughout the Years

Hernando Alice Geraldine S^{1,2*}, Susana F. Baldia^{1,3}, Paciente A. Cordero Jr.^{1,4}

¹The Graduate School, University of Santo Tomas, Sampaloc, Manila, PHILIPPINES. ²College of Arts and Sciences, Mariano Marcos State University, City of Batac, Ilocos Norte, PHILIPPINES. ³Research Center for the Natural and Applied Sciences, University of Santo Tomas, Sampaloc, Manila, PHILIPPINES. ⁴Eastern Visayas State University, Burauen Campus, Burauen, Leyte, PHILIPPINES.

Submission Date: 02-05-2020; Revision Date: 20-06-2020; Accepted Date: 25-06-2020

ABSTRACT

Microalgae or commonly known as phytoplankton are the earliest and oldest forms of life on this living planet and considered as tiniest plant in the archipelago. These organisms display different colorations in water such as green, blue-green and brown. At present, phytoplankton are gaining much attention for feed, food and other important products in various fields and industries yet they are thought of as the most poorly studied group of aquatic organisms. Noteworthy, some of the microalgae which exist already in the commercial market are Spirulina, Chlorella, Haematococcus and Chaetoceros. Because of this, industrial microalga is a field that needs to be exploited primarily to produce promising high-valued chemicals for nutraceuticals, functional food and living feed and other feed additives. Additionally, these creatures were visualized as the "food for the future" because of the many applications that it possess. Historically, the first microalgae to be commercialized for food industry was the Nostoc species over many decades and consumed in China, Taiwan, Japan and other Southeast Asian Nations. Briefly, Nostoc species is a blue-green alga and sometimes called freshwater grapes which are in a jelly-like ball surrounded by thick mucilage. Thus, this review article aims to present the background antecedents of how phytoplankton discovered and existed in the Philippines for many decades and to discuss various phytoplankton with their biological components necessary for agriculture, fisheries, industrial, pharmaceutical, medical and other fields of endeavor.

Key words: Biological, Chemical, Constituents, History, Minute.

INTRODUCTION

Algae are polyphyletic assemblage of organisms that are diversely found in nature which can be in the form of one-celled, multi-celled, filamentous, made up of simple reproductive structures and can live in harsh, extreme conditions. These species are divided into either macro or micro algae with microalgae or commonly known as seaweeds which can grow up to 50 meters long and above (e.g. Genus *Macrocystis*). Microalgae or phytoplankton

SCAN QR CODE TO VIEW ONLINE	
	www.ajbls.com
	DOI : 10.5530/ajbls.2020.9.37

are tiny, floating and self-feeder organisms which are found and evenly distributed in various bodies of water in which the species can synthesize their own food. Most of the species are unicellular but some are colonial which are usually structured uniformly. Microalgae are segregated into hierarchical position and eventually identified by certain cell characteristics such as to their cell shape, cell size, cell wall flexibility, presence of chloroplasts and definite pigments and the number of the flagella.^[1]

BODY

Historical antecedents

The 1853 to 1862 algal collections by Bailey and Harvey isolated the first Philippine Diatom. Likewise, the work

Correspondence: Prof. Alice Geraldine S. Hernando, ¹The Graduate School, University of Santo Tomas, Sampaloc, Manila, PHILIPPINES. ²Department of Biological Sciences, College of Arts and Sciences, Mariano Marcos State University, City of Batac, Ilocos Norte, PHILIPPINES.

Phone no: +63 09085952400 Email: alicegeraldin_ hernando@yahoo.com of O'Meara in 1872 isolated the Genus Navicula from the Sulu Archipelago.^[2,3] In the 1900's, Mann in 1925 published a book on marine diatoms of the Philippines and this was followed by several studies on both marine and freshwater diatoms.^[2] Also, the diversity of phytoplankton at Lake Bato in Camarines sur contained 61 genera was studied.^[4] Moreover, the abundance of phytoplankton in Manila Bay,^[5] subsequently the presence of microalgae in the ponds and pools in Manila and its vicinity^[6] and later the diversity of phytoplankton in Laguna de Bay^[7] were surveyed and conducted. Diatoms and dinoflagellates in Northwestern Luzon^[8] and Northwestern Iloilo^[9] were also examined and identified. Recently, microalgae distributed in the seven (7) lakes of San Pablo City and the Crocodile Lake in Los Baños were isolated and cataloged.^[10] In addition, four (4) phyla of microalgae was also noted and studied in Lake Paoay and Lake Mohicap,^[11,12] and.^[13] More recent, microalgae including cyanobacteria found on the walls of the various buildings of the University of the Philippines, Los Baños, Laguna were isolated and identified. The morphological and cytological structures of the Chlorophycea and Bacillarophycea micro algal species were documented.^[14] Likewise, micro algal species were evaluated at the Fish Cage Belt at the Magat Reservoir, Philippines. Phytoplanktons such as Scenedesmus spp. Nitzschia spp. and Closterium spp. dominated the area. Also, water quality assessment such as the temperature, pH and dissolved oxygen was gathered on the said reservoir.^[15]

The following morphological features and taxonomic identification are according to the studies conducted on the diversity of various ecosystems and places in the Philippines. Chlorophytes or commonly known as the green algae are typically unicellular but some are multicellular or even colonial in nature. They are typically found in freshwater, but some species are seen in the sea. These microalgae are composed of membrane-bound chloroplasts, nuclei and flagella bearing 2-4 cells. They are green in color due to the presence of chlorophyll a and b embedded in the chloroplast. Many species can duplicate asexually by fission and sexually by iso- or anigamous. These species can invade various aquatic ecosystems such as freshwater, marine habitats and terrestrial areas. Freshwater green algae includes charophytes; marine microalgae includes chlorophytes and terrestrial species are mostly trebouxiophytes. Cyanophytes or the bluegreen algae are well-known for its affinity to bacteria and typical called the blue-green algae. The species are prokaryotic and their sizes ranged from 0.2-2 um. The cells do not have nuclei and lack double membranes as well as chromosomes, but capable of duplication.

However, the cell walls are double-membrane bounded organelles making them rigid. Their bodies are made up of large vacuole for buoyancy control. Just like any other microalgae, the various species can also be colonial and filamentous in habit. They were identified as components of the Pico plankton particularly in the temperate areas like the country. Finally, the Bacillariophytes or the diatoms which are ubiquitous, autotrophic, unicellular, filamentous or colonial and free swimming organisms. They are mostly colorless, but photosynthetic in nature. Scientists observed that this phylum is also facultative heterotrophic. Most of the species have a unique characteristic which is the presence of siliceous skeleton or frustule. The siliceous box of these organisms are composed of cytoplasm, nucleus and vacuole. Moreover, the body itself is also made up of chloroplasts which are yellow brown in color because of the xanthophyll such as fucoxanthin pigment dominating over that of the chlorophyll a and c. Species of this phylum are important in lakes because they are source of food for the zooplankton.

Biological components

Phytoplankton have an amazing number of biological compounds necessary for industrial and biotechnological purposes. The following are some of the discovered and reported compounds excreted by the algal flora. Micro algal species are made up of various Essential Fatty Acids (EFA) or EFA for a healthy diet in order to grow, develop and meet the dietary requirements.^[16] These compounds are the structural and physiological components of lot of lipids and different phytoplankton have varied EFA components depending on the biological functioning of the species.^[17] Phytoplankton have also Sterols which are one of the vital constituents of these organisms and tagged as the blueprint of organic matter in the aquatic habitat.^[18] Depending on the functionality of the micro algal species, it varies in terms of sterol metabolic component as well as biological pattern. Cholesterol is the most abundant sterol having variety of sources and functions.^[19] Generally, carotenoids have become prominent value added-product from algae particularly in the US with a market value of 1.2 billion and gaining more because of its health benefits circulating in the world^[20] such as for food supplements, vitamin increments and feed additives for both poultry and aquaculture.^[21] This pigment has 600 various types derived from 40 carbon chain. The most active type is the B-carotene which acts as pro-vitamin A, additive in multivitamin supplements and promote an antioxidant capacity. Dunaliella spp. and Spirulina spp. contain this compound making them ideal as food supplements

and feed nutritive value.^[22] Astaxanthin is a carotenoid with a red-colored pigment and considered to be the super vitamin E because of its antioxidant capacity^[23-25] and has a defense mechanism to certain organisms.^[26] It is also the pigment used to protect organisms from strong ultra-violet rays and DNA alterations in skin fibroblast.^[27] This pigment has been examined in some aquatic organisms including microalgae such as Dunaliella salina and Spirulina maxima and found in crustaceans such as shrimps and fishes like salmon and trout. It is costly in salmon farming containing 15% of the total production^[28] hence, the most important carotenoid in salmons and in trout.^[29] Another carotenoid pigment is Lutein which is known to delay the occurrence of cataract, decreasing the vision particularly the agerelated diseases and prevent from blindness by making the eyes healthy and free from any danger. This pigment is also responsible in the natural coloration of foods and cosmetics.^[30-32] The market value of this pigment is also increasing just like the other carotenoids from 139 million dollar to 233 million dollar. Chlorella sp., Muriellopsis sp. and Scenedesmus sp. are the first microalgae to be discovered with lutein.^[21,32] Beta-Carotene is considered as one of the most significant type of carotenoids, this pigment has the same activity with the Vitamin A or retinol for human body.^[33] This is also used as coloring agent and in pharmaceutical and cosmetic industry.^[17,20] This compound has proven to be a strong antioxidant which scavenge free radicals and fight cancers and other diseases.^[33] The market value of this pigment increased from 261 million dollar as of 2010 to 334 million dollar by 2018. Dunaliella strains are the largest group of microalgae to produce large quantities of beta-carotene.^[20] Phycobiliproteins are water-soluble compounds and light capturing proteins excreted by many micro algal species.^[34] The metabolite is in 3 forms according to ultraviolet absorption such as phycocyanin, phycoerythrin and allophycocyanin. It is a component of the natural dyes made by pharmaceutical and cosmetic industries, used as fluorescent tags and immunological diagnostics agents. The compound was first observed in the cyanobacteria Spirulina sp.[20] Recently, it was observed in 2 other cyanobacteria namely Oscillatoria sp. and Scytonema sp.^[35] It has a market value of 5-10 million and above.^[36] Another is the Polyunsaturated fatty acids or PUFAs which are polymeric and have several types namely docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA), arachidonic acid (AA) and gamma linolenic acid (GHA). These are also nutritional and functional foods or ingredients in the animal diet including fishes. It has a market value in the US of about 7.2 billion dollar in 2011 and reached 13 billion dollar in 2017.^[20] The sources of these fatty acids are from fishes but poorly act as additives having huge stability extracted from algal oils depending on physical parameters such as changing culture conditions like temperature, light intensity and concentration of nitrogen.[37-39] Various marine and freshwater marine algae are made up of the different PUFAs. Also, these compounds are incorporated in infant and full term food formulas for healthier growth, develop the infant brain and to fight disease causing microorganisms.^[40,41] In addition, Beta- Glucan are also compound which are known traditionally as Zymosan in Norway in the 1940's but finally named beta glucan in Italy in the 1960's because it was found in the baker's yeast particularly in its cell wall.^[42] This pigment is important in human health as well as in fish's lifestyle for bacterial attack and infection.^[43] Chlorella spp. are the major sources of this metabolic compound.[44] Finally, Mycosporinelike amino acids or MAA's which are compounds that are prominently synthesized by marine and some freshwater microalgae. However, the most primitive maker and producers of these secondary metabolites are cyanobacteria.^[45] The unique characteristic of these compounds includes high ultraviolet absorption.[46]

Phytoplankton species have been out in the commercial market for several years. A number of them have been very successful in invading the regional, national and international markets. Four (4) phytoplankton are highlighted and their importance as natural food, supplement and pharmaceutical products were discussed. Spirulina spp. or Arthrospira exist as freefloating filaments with cylindrical multicellular trichomes, can tolerate high pH and can thrive in alkaline hot lakes.^[47,48] It can be found in freshwater and seawater habitats both in tropical and temperate regions and can live either in alkaline and salty waters.^[49-51] It is excellent in protein for its nutritive value^[39,52,53] with an amino acid component of 62-68% more than enough compared to the components of beef, rich in vitamins A and B, contains beta carotene ten times than carrots, made up of essential fatty acids and linoleic acid, as well as linolenic acid, eicosapentaenoic acid, docosahexaenoic acid and arachidonic acid and a polysaccharide named Immulina where it improves the immune system of animals.^[54-56] This cyanobacterium is one of the most useful species for commercialization for healthy food, animal supplement and nutritional products with health benefits.^[57-59] It is also the first micro algal species that dominated the commercial industry for many years after the establishment of Nostocsp. as the first commercialized microalgae in Asia with many therapeutic and toxicological studies that have been conducted and

reported.^[17] It was emphasized as food for tilapia;^[60] rockfish^[61] along with other microalgae species namely Chlorella spp., Isochrysis spp., Scenedesmus spp., Pavlova spp., Dunaliella spp. Chaetoceros spp. and Tetraselmis spp.^[62] Consequently, Spirulina platensis together with Oscillatoria sp. and Nostoc muscorum with concentrations ranging from 82-100%^[63] as well as Dunaliella sp., Tetraselmis sp. and Chlorella sp. in their acetone extract^[64] were evaluated and proved to be effective for their antifungal activities. These are due to the presence of long chain of fatty acids such as palmitoleic, oleic, linoleic and linolenic acid as an attribution of their antifungal activities^[65,66] In addition, Spirulina platensis along with Anabaena oryzae, Nostoc muscorum and Phormedium fragile as well as the Genus Oscillatoria sp. were also proven to kill nematodes hence, acted as nematicidal agent.^[67] Correspondingly, Genus Spirulina is used as a feeding supplement and functional food to intestinal flora like Lactobacillus and Bifidus.^[68] In humans, consuming the Spirulina products lowered the cholesterol level of the human serum.^[69] As to the impairment of the bone marrow, the metabolites phycocyanin stimulate hematopoiesis, thus, regulating hormones needed to rehabilitate the damage. Spirulina also treats hypocholesterolemia, hyperlipidaemia and Artherosclerosis.^[70] The effectivity of this micro algal in food and feed industry is because of the incorporation of the whole biomass of the algae providing green colorant thus, escalating the nutritional values.^[71] Another is *Chlorella* spp. which are unicellular or colonial microscopic organisms containing the two (2) chlorophylls a and b. These organisms contain 45% of protein, 20% of lipid, 20% of carbohydrates and other vitamins and minerals,^[48,72] They are easy to cultivate because it requires simple conditions such as the presence of water, carbon dioxide, light and some minerals only.[73] The largest producer of Chlorella worldwide is the Taiwan Chlorella Manufacturing and Co. found in Taipei, Taiwan. The company generate approximately 400 tons of dried micro algal biomass per year.^[74,75] It is also known and prominent in ASEAN countries and to the United States with a total sale of 38 billion dollars and above annually.^[76] It was first cultivated in 1910 for aquaculture industry as animal feed in Berlin, Germany.^[77] Most of the commercialized Chlorella is in the form of capsule and caplets.^[17] In Japan, the Chlorella ellipsoidea is commercialized and used as a dietary aid for powdered green tea, soup, noodles, rolls, cookies and ice cream.^[75] Likewise, it is also sold in the market of Japan as colorant with a name "Lina-blue".[78] This genus is also a component of the healthy diet of humans and other forms of animals. The species is also used for

medication such as treating anemia, gastric orders, constipation, wounds and hypertension. In addition, this algal species proved to escalate the concentration of hemoglobin, lower down sugar levels and an agent for hepatoprotective activity. This species was found to have the metabolite glucan which serve as a free radical scavenger reducing the blood lipids and increase the energy and wealth of most leprosy patients.^[74,75,79] In specific, the species Chlorella zofingiensis produced also the metabolite astaxanthin under conditions of limited nitrogen and no presence of light for pharmaceutical purposes.^[80] This species also excrete other metabolites useful for beauty and personal skin products to promote glowing and natural beauty.^[81] C. pyrenoidosa and C. vulgaris are also made up of amino acids such as glycine and proline which are rich in biological fragments.^[82] In addition, Chlorella vulgaris.^[83,84] Dunaliella sp., Synechocystis sp. and Tetraselmis sp.^[85,86] together with other micro algal species namely Nostoc muscorum, N. humifusum, Anabaena flos aquae, A. oryzae, Spirulina platensis, Phormedium fragile, Wollea saccata and an unknown species of the Genus Oscillatoria were identified with strong antioxidant activity in their aqueous forms. The study was done through the DPPH assay and species have percentage of antioxidative capacity of 30.1%-72.4% with compounds such as phenols, terpenoids, alkaloids; phycobiliprotein pigments and flavonoids which are effective as antioxidant and tumoristatic activity.[87] In other study, Chlorella vulgaris together with Dictyochloropsis splendida and Scenedesmus obliquus were also examined for their antioxidant capacity yielding high presence of carotenoids and phenols^[88,89] and together with Nannochloropsis oculata and Tetraselmis tetrathele were also investigated to have an inhibitory activity against lipid peroxidation to linoleic acid so having an antioxidant activity.^[90] Meanwhile, Scenedesmus spp. are green algae can tolerate various physiological conditions such as temperature from 15 to 40°C; optimal temperature of 30°C^[91] and pH of neutral or 7.^[92] For this, Scenedesmus is conceived to be viable as source of biological applications.^[93] This is also due to the huge amount of nutritional components such as protein, lipids, carbohydrates and other trace elements.^[94] Likewise, it is ideal as food constituents and animal feed and additives. ^[95] These organisms are identified as nutritional food source commonly added in desserts, fruit pudding and soup but the commercialization is still limited.^[75] Although in few studies, the extracts of this species together with Chlorella sp. are used for human consumption^[96] in which this species generally produced polysaccharides that act as oxidative agent^[97] and extracellular enzymes or exoenzymes to degrade

environmental pollutants.^[98] In specific, Scenedesmus costatum have proven to excrete a long chain of fatty acids causing the species an antibacterial agent against aquatic bacteria along with diatom, Thalassiosira rotula and S. costatum which are composed of antibacterial compound oxylipins^[17] and Chaetoceros muelleri and Dunaliella salina^[99] and Chlorella spp. have antibacterial activities with isolated mixtures of fatty acids and chlorellin inhibiting both gram positive and gram negative microorganisms particularly bacteria.[100-102] Similarly, this microalgae can also treat and manage wastewater, hence, it has a dual role such as treatment of wastewater and producer of biomass.^[103,104] Finally, Dunaliella spp. are eukaryotic photosynthetic organisms which are made up of two or more flagella for locomotion.^[105] It is capable of tolerating high demand of salt concentrations in a given habitat. Thus, halotolerant microalgae which naturally occurs in salted lakes contains up to 10% of beta carotene under stressful conditions such as low nutrient and salt.[106] Dunaliella spp. can be found in various habitats in oceans, seas, salt lagoon and salt marshes. These algae are mainly the source of carotene or carotenoids in the commercial market. In specific, Dunalliela salina has an estimated market size of 10-100 tonnes per year and has a market prize of 750 per kg. The main producers are Australia, Israel, USA, China.^[107] This is cultivated in large scale by the open pond system and protected by the carotenoid pigment against high level of climatic conditions.^[108] This algal species is ideal for biofuel production due to its intracellular lipid content of 67% of the total dry weight.^[109] It is also used in molecular farming or commonly known as biopharming where it expressed the hepatitis B surface antigene gene that makes the vaccine and antibody production more uncostly.[110,111]

CONCLUSION

Therefore, these phytoplankton are diversified in terms of characteristics, components and utilization. There are still a lot to discover on these minute, little organisms with vast array of biological importance. Moreover, utilization of the phytoplankton itself and their components are unexplored and dearth of micro algal studies which are deemed necessary to boost and hone this another small but gigantic era of biological field of study. Furthermore, discovery of experts on this endeavor is vital to boom this phytoplankton research and development.

ACKNOWLEDGEMENT

This is to acknowledge the Commission on Higher Education (CHED) and Mariano Marcos State University (MMSU), for the support in this study.

CONFLICT OF INTEREST

The authors declare no Conflict of interest.

REFERENCES

- Boney AD. Phytoplankton. New Studies in Biology. 2nd Edition. ISBN 0-7131-2978-6.
- JrCordero P A. Philippine Algology: Its Beginnings and Development. Leyte-Samar Studies. 1972;6(1):16-37.
- Relon ML. Microalgae in Talin Bay. Manila: De La Salle University. Press. ISBN 9-7155-5410-5.
- Perez SA. Planktonic composition of Lake Bato, Camarines Sur. M. Sc. Thesis, Philippine Women's University Graduate School, Manila. 1969;210.
- Tiews KS, Bravo A, Ronquillo A. On the Food and Feeding Habits of some Philippine shrimp in Manila Bay and San Miguel Bay. Philippine Journal of Science. 1976;14(2):204-11.
- Martinez MR, Pantastico JB. Some common algae found in ponds and pools. Philippine Biota. 1976;10(3):81-6.
- Pantastico JB. Taxonomy of the Freshwater Algae of Laguna de Bay and Vicinity. National Research Council of the Philippines. Bull. 1977;261.
- Relon ML, Cordero PA. Taxonomy of the phytoplankton flora in Northwestern Luzon, Philippines with notes on their Ecology. Ph.D. Dissertation. University of Santo Tomas, Espana, Manila. 1985;276.
- Escuadra CT. Morpho-Taxonomy of phytoplankton in Northeastern Iloilo specifically in the vicinity of Estancia and Carles. M. Sc. Thesis. University of Santo Tomas, Espana, Manila. 1983;234.
- Zafaralla MT. Microalgae of the Seven Lakes of San Pablo City and Crocodile Lake of Los Baños. University of the Philippines Los Baños, Philippine Council for Agriculture, Aquatic and Natural Resources Research and Development, Department of Science and Technology. Philippines. 2014
- Villaroman KMD, Baldia SF. Phytoplankton Community Structure of Paoay Lake (Ilocos Norte, Philippines). M. Sc. Thesis. Graduate School. University of Santo Tomas, Manila. 2012.
- Cordero CS, Baldia SF. Biodiversity of Phytoplankton Community in Lake Mohicap, San Pablo City, Laguna, Philippines. M. Sc. Thesis. Graduate School, University of Santo Tomas, Manila. 2015.
- Sambitan KA, Papa RD, Baldia SF. Phytoplankton community structure of Lake Paoay and Lskr Mohicap with notes on the first record of Ceratium (Dinophyta) in Lake Paoay. Acta Manilana. 2015;63:51-60.
- Arguelles EDLR. Morphotaxonomic Account of Epilithic Microalgae and Cyanobacteria in Los Baños, Laguna (Philippines). Research Gate. 2016;17. Print ISSN 2244-1573 • Online ISSN 2244-1581.
- Baleta FN, Bolaños JM. Phytoplankton identification and water quality monitoring along the fish-cage belt at Magat dam reservoir, Philippines. International Journal of Fisheries and Aquatic Studies. 2016;4(3):254-60.
- Valeem EE, Shameel M. Fatty acid composition of blue-green algae of Sindh, Pakistan. Int Journal of Phycology and Phycochemistry. 2005;1(1):83-92.
- 17. Mostafa SSM. Microalgal Biotechnology: Prospects and Applications. Plant Science. 2012. INTECH. Chapter 12. http://dx.doi.org/10.5772/53649.
- Ponomarenko LP, Stonik IV, Aizdaicher NA, Orlova TY, Popvskaya GI, Pomazkina GV, et al. Sterols of marine microalgae Pyramimonas cf. cordata (Prasinophyta), Atteya ussurensis sp. nov. (Bacollariophyta) and a spring diaom bloom form Lake Baikal. Comp Biochem Physiol B. 2004;138(1):65-70.
- Froehner S, Martins RF, Errera MR. Assessment of fecal sterols in Barigui River sediments in Curitiba, Brazil. Environment Monitoring Assessment. 2009;157(1-4):591 Doi. 10.1007/s10661-008-0559-0.
- Yaakob Z, Ali E, Zainal A, Mohamad M, Takriff MS. An overview: biomolecules from microalgae for animal feed and aquaculture. Journal of Biological Research. 2014;21(1):6.

- Del-Campo JA, Garcia-Gonzales M, Guerrero MG. Outdoor cultivation of microalgae for carotenoid production: current state and perspective. Appl Microbiol Biotechnol. 2007;74(6):1163-74.
- Becker EW. Microalgae for Human and Animal Nutrition. Handbook of Microalgal Culture: Applied Phycology and Biotechnology. John Wiley and Sons. 2013.
- Naguib YMA. Antioxidant activities of astaxanthin and related carotenoids. J Ag Food Chem. 2000;48(4):1150-4.
- Kupcinskas L, Lafolie P, Lignell A, Kudelis G, Jonaitis L, Adamonis K, et al. Efficacy of the natural antioxidant astaxanthin in the treatment of functional dyspepsia in patients with or without *Helicobacter pylori* infection: A prospective, randomized, double blind and placebo-controlled study. Phytomedicine. 2008;15(6-7):391-9.
- Nakagawa K, Kiko T, Miyazawa T, Burdeos GC, Kimura F, Satoh A, *et al.* Antioxidant effect of astaxanthin on phospholipid peroxidation in human erythrocytes. Bri J Nutr. 2011;105(11):1563-71.
- Yuan JP, Peng J, Yin K, Wang JH. Potential health-promoting effects of astaxanthin: A high-value carotenoid mostly from microalgae. Mol Nutr Food Res. 2011;55(1):150-65.
- Lyons NM, O'Brien NM. Modulatory effect of an algal extract containing astaxanthin on UVA-irradiated cells in culture. J Dermatol Sci. 2002;30(1):73-84.
- Mazhar D, Ang R, Waxman J. COX inhibitors and breast cancer. Br J Cancer. 2006;94(3):346-50.
- Tolasa S, Cakli S, Ostermeyer U. Determination of astaxanthin and canthaxanthin in salmond. Eur Food Res Technol. 2005;221(6):787-91.
- Shi XM, Chen F, Yuan JP, Chen H. Heterotrophic production of lutein by selected *Chlorella* strains. J Appl Phycol. 1997;9(5):445-50.
- Zhang XW, Shi XM, Chen F. A kinetic model for lutein production by the green algae *Chlorella protothecoides* in heterotrophic culture. J Ind Microbiol Biot. 1999;23(6):503-7.
- 32. Wu ZY, Shi CL, Shi XM. Modeling of lutein production by heterotrophic *Chlorella* in batch and fed-batch cultures. World J Microb Biot. 2007;23(9):1233-8.
- Chen BH, Chuang JR, Lin JH, Chiu CP. Quantification of pro-vitamin A compounds in Chinese vegetables by high-performance Liquid Chromatography. J Food Prot. 1993;56(1):51-4.
- Liu L, Pohnert G, Wei D. Extracellular Metabolites from Industrial Microalgae and their Biotechnological Potential: A Review. Marine Drugs MDPI. 2016;14(10):191. doi.10.3390/md14100191.
- Karseno HK, Bamba T, Dwi S, Mahakhant A, Yoshikawa T, Hirata K. Extracellular phycoerythrin-like protein released by freshwater cyanobacteria Oscillatoria and Scytonema sp. Biotechnol Lett. 2009;31(7):999-1003.
- Sekar S, Chandramohan M. Phycobiliproteins as a commodity: Trends in applied research, patents and commercialization. J Appl Phycol. 2008;20(2):113-36.
- 37. Reitan KI, Rainuzzo JR, Olsen Y. Effect of nutrient limitation on fatty acid and lipid content of marine microalgae. J Phycol. 1994;30(6):972-9.
- Liang Y, Beardall J, Heraud P. Effects of nitrogen source and UV radiation on the growth, chlorophyll fluorescence and fatty acid composition of *Phaeodactylum tricornutum* and *Chaetoceros muelleri* (Baccilariophyceae). J. Photochem Photobiol Biol. 2006;82(3):161-72.
- Spolaore P, Joannis-Cassan C, Duran E, Isambert A. Commercial applications of microalgae. J Biosci Bioeng. 2006;101(2):87-96.
- Patil V, Kallqvist T, Olsen E, Vogt G, Gislerod HR. Fatty acid composition of 12 microalgae for possible use in aquaculture feed. Aquacult Int. 2007;15(1):1-9.
- Kousoulaki K, Ostbye TKK, Krasnov A, Torgensen JS, Morkore T, *et al.* Metabolism, Health and fillet nutritional quality in Atlantic salmon, *Salmo salar* fed diets containing n-3-ric microalgae. J Nutri Sci. 2015;4:e24.
- 42. Riggi SJ, DiLuzio NR. Identification of a reticuloendothelial stimulating agent in zymosan. Am J Physiol. 1961;200(1):297-300.
- Sahoo PK, Mukherjee SC. Effects of dietary beta 1,3 glucan on immune system responses and disease resistance of healthy and aflatoxin B1induced immune compromised rohu (Laheo rohita Hamilton). Fish Shellfish Immun. 2001;11:683-95.
- Iwamoto H. Industrial production of microalgal cell-mass and secondary products-major industrial species *Chlorella*. In Handbook of Micro Algal Culture Biotechnology and Applied Phycology. Blackwell Publishing Ltd. 2004;255-63.
- Klisch M, Hader D. Mycosporine-Like Amino Acids and Marine Toxins- The Common and the Different. Marine Drugs. 2008;6(2):147-63.

- Takano S, Nakanishi A, Uemura D, Hirata Y. Isolation and structure of a 334 nm UV absorbing Substance, porphyra-334 from the red algae *Porphyra tenera* Kjellman. Chem Lett. 1979;8(4):419-20.
- Habib MAB, Parvin M, Huntington TC, Hasan MR. A review on Culture, Production and Use of *Spirulina* as Food for Humans and Feeds for Domestic Animals and Fish. Food and Agriculture Organization of the United Nations. Rome, Italy. 2008;28-32
- Nicoletti M. Microalgae Nutraceuticals: A Review. MDPI Foods. 2016;5(3):54. Doi.10.3390/foods5030054.
- 49. Farrar WV. Tecuitlatl: A glimpse of Aztec food technology. Nature. 1966;211:341-2.
- 50. Ciferri O. Spirulina, the edible microorganism. Microbiol Rev. 1983;47(4):551-78.
- 51. Shao-Chen GUAN. Study on Seawater-acclimation *Spirulina*. Journal of Anhui Agricultural Sciences. 2012;6:132.
- Campanella L, Crescentini G, Avino P. Chemical composition and nutritional evaluation of some natural and commercial food products based on *Spirulina*. Analysis. 1999;27(6):533-40.
- 53. Campanella L, Russo MV, Avino P. Free and total amino acid composition in blue-green algae. Ann Chim. 2002;92(4):343-52.
- Juarez-Oropeza MA, Mascher D, Torres-Duran PV, Farias JM, Paredes-Carbajal MC. Effects of *Spirulina* on vascular reactivity. J Med Food. 2009;12(1):15-20.
- Capelli B, Cysewski GR. Potential Health Benefits of *Spirulina* Microalgae. A Review of the Existing Literature. Cyanotech Corporation: Kailua-Kona, Hi, USA. 2010;9(2):19-26.
- Pugh NSA, Ross HN, ElSohly MA, ElSohly DSP. Isolation of three high molecular weight polysaccharide preparations with potent immunostimulatory activity from Spirulina platensis, aphanizomenon flos-aquae and Chlorella pyrenoidosa. Planta Med. 2001;67(8):737-42.
- Borowitzka MA. Commercial production of microalgae: ponds, tanks, tubes and fermenters. Journal of Biotechnology. 1999;70:313-21.
- Kang HK, Salim HM, Akter N, Kim DW, Kim JH, Bang HT, et al. Effect of various forms of dietary Chlorella supplementation on growth performance, immune characteristics and intestinal microflora population of broiler chickens. Poultry Science Association, Inc. Research Report. 2013;22(1):100-8.
- Cho WY, Kang DH, Lee HY. Enhancement of immune activation activities of Spirulina maxima grown in deep-sea water. Int J Mol Sci. 2013;14(6):12205-21.
- 60. Tartiel M, et al. Partial replacement of fish meal with dried microalga (Chlorella spp. and Scenedesmus spp) in Nile tilapia Oreochromis niloticus diets. 8th International Symposium on Tilapia in Aquaculture. Central Laboratory for Aquaculture Research, Agricultural Research Center, Ministry of Agriculture, Egypt.2008;801-11.
- Bai SC, Ko JW, Kim KW, Kim SK. Effects of *Chlorella* powder as a fed additive on growth performance in juvenile Korean rockfish, *Sepbastes schlegeli* (Hilgendorf). Research Gate. Agricultural Research. 2001;32(1):92-8.
- Hejazi MA, Kleinegris D, Wijffels RH. Mechanism of extraction of betacarotene from microalga *Dunaliellea salina* in two-phase bioreactors. Biotechnol Bioeng. 2004;88(5):593-600.
- 63. Hussein MY, Abd EA, Azza AM, Mostafa SSM. Bioactivity of algal extracellular byproducts on cercospora leaf spot disease, growth performance and quality of sugar beet. The 4th Conference on Recent Technologies in Agriculture: Challenges of Agriculture Modernization, Nov. 3th-5th, Special Edition of Bull Fac Agric Cairo Univ. 2009;1:119-29.
- Rajendran N, Selvan K, Piriya SP, Logeswari V, Kathiresan E, Tamilselvi A, *et al.* Phytochemicals, Antimicrobial and Antioxidant Screening from Five Different Marine Microalgae. Journal of Chemical and Pharmaceutical Sciences. National Conference on Plant Metabolomics. 2014. ISSN: 0974-2115.
- Agoramoorthy G, Chandrasekaran M, Venkatesalu V, Hsum MJ. Antibacterial and antifungal activities of fatty acid methyl esters of the blind your eye mangrove from India. Braz J Microbiol. 2007;38(4):739-42.
- Plaza M, Santoyo S, Jaime L, Reina GGB, Herrero M, Senorans FJ, et al. Screening for bioactive compounds from algae. J Pharm Biomed Anal. 2010;51(2):450-5.
- Dhanam M, Kumar AC, Sowajanya AS. *Microcoleu vaginatus* (Oscillatoriacea), a blue-green algae (*Cyanobacterium*) parasitizing plant and soil nematodes. Indian Journal of Nematology. 1994;24:125-132.
- Priyadarshani I, Rath B. Commercial and industrial applications of microalgae: A review. J Algal Biomass Utln. 2012;3(4):89-100.

- DeGonzales RC, Miranda-Zamora R, Diaz-Zagoya JC, Juarez OMA. Preventive effect of *Spirulina maxima* on the fatty liver induced by a fructose rich diet in rat, a preliminary report. Life Sci. 1993;53(1):57-61.
- Eussen S, Klungel O, Garssen J, Verhagen H, Kranen HV, Loveren HV, et al. Support of drug therapy using functional foods and dietary supplements: focus on statin therapy. Br J Nutr. 2010;103(9):1260-77.
- Danesi EDG, Rangel-Yagui CO, DeCarvalho JCM, Sato S. An investigation of effect of replacing nitrate by urea in the growth and production of chlorophyll by *Spirulina platensis*. Biomass and Bioenergy. 2002;23(4):261-9.
- Herrador M. The Microalgae/Biomass Industry in Japan: An Assessment of Cooperation and Business Potential with European Companies. Final Report. 2016;1-170.
- Bertoldi FC, Sant'anna E, Oliveira JLB. Chlorophyll content and mineral profile in the microalgae *Chlorella vulgaris* cultivated in hydroponic wastewater. Cienc Rural. 2008;38(1):54-8.
- Gouveia L, Baptista AP, Ray-mundo SI, Bandarra NAM. Microalgae in novel food products. Food Chemistry Research Developments. Nova Science Publishers, New York. 2008;269-74.
- Sathasivam R, Radhakrishnan R, Hashem A, Abd AE. Microalgae metabolites: A rich source for food and medicine. Saudi Journal of Biological Sources. Science Direct. 2017. https://doi.org/10.1016/j.sjbs.2017.11.003.
- Yamaguchi K. Recent advances in microalgal bioscience in Japan, with special reference to utilization of biomass and metabolites: A review. J Appl Phycol. 1997;8:487-502.
- Preisig HR, Andersen RA. Historical review of algal culturing techniques. In Algal Culturing Techniques. Elsevier Academic Press. 2009;1-12.
- Kovac DJ, Simeunovic JB, Babic OB, Misan AC, Milovanovic IL. Algae in Food and Feed: Review Article. Food and Feed Research. 2013;40(1):21-31.
- Grima EM, Belarbi EH, Fernandez FGA, Medina AR, Chisti Y. Recovery of microalgal biomass and metabolites: Process options and economics. Biotechnology Advances. 2003;20(7-8):491-515.
- Pelah D, Sintov A, Cohen E. The effect of salt stress on the production of canthaxanthin and astaxanthin by *Chlorella zofingiensis* grown under limited light intensity. World J Microbio Biotechnol. 2004;20(5):483-6.
- Jaspars M, DePascale D, Andersen JH, Reyes F, Crawford AD, Ianora A. The Marine biodiscovery pipeline and ocean medicines of tomorrow. J Mar Biol Assoc UK. 2016;96(1):151-8.
- Selvaraj G, Kaliamurthi S, Cakmak ZE, Cakmak T. Computational screening of microalgae and cyanobacteria RuBisCO as potential precursor for bioactive peptides. Peer J Preprints. https://doi.org/10.7287/peerj.preprints. 2017;1-19.
- Mendiola JA, Rodriguez-Meizoso I, Senorans FJ, Reglero G, Cifuentes A, Ibanez E. Antioxidants in Plant Foods and Microalgae extracted using compresses fluids. Electron J Environ Agric Food Chem. 2008;7:3301-9.
- Shanab SMM, Mostafa SSM, Shalaby EA, Mahmoud GI. Aqueous extracts of microalgae exhibit antioxidant and anticancer activities. Asian Pacific Journal of Tropical Biomedicine.2012;12(8):608-15.
- Herrero M, Ibanez E, Cifuentes A, Reglero G, Santoyo S. *Dunaliella salina* microalgae pressurized liquid extracts as potential antimicrobials. J Food Prot. 2006;69(10):2471-7.
- Rajendran N, Selvan K, Piriya SP, Logeswari V, Kathiresan E, Tamilselvi A, et al. Phytochemicals, Antimicrobial and Antioxidant Screening from Five Different Marine Microalgae. Journal of Chemical and Pharmaceutical Sciences. National Conference on Plant Metabolomics. 2014. ISSN: 0974-2115.
- Li HB, Cheng KW, Wong CC, Fan KW, Chen F, Jiang Y. Evaluation of antioxidant capacity and total phenolic content of different fractions of selected microalgae. Food Chem. 2007;102(3):771-6.
- Natrah FMI, Yusoff FM, Shariff M, Abas F, Mariana NS. Screening of Malaysioan indigenous microalgae for antioxidant properties and nutritional value. J Appl Phycol. 2007;19(6):711-8.
- Ali HEA, Shanab SMM, Abo-State MAM, Shalaby EAA, Demerdash UMN, Abdullah MA. Screening of Microalgae for Antioxidant Activities, Carotenoids and Phenolic Contents. Applied Mechanics and Materials. 2014;625:156-9.
- 90. Pumas P, Pumas C. Proximate Compositon, Total Phenolics Content and Antioxidant activities of Microalgal Residue from Biodiesel Production: Maejo

International. Journal of Sciene and Technology. 2014;8(02):122-8. ISSN 1905-7873.

- Martinez ME, Jimenez JM, Yousufi F. Influence of phosphorus concentration and temperature on growth and phosphorus uptake by the microalga *Scenedesmus obliquus*. BioResource Technology. 1999;67(3):233-40.
- Hodaifa G, et al. Biomass production of Scenedesmus obliquus from mixtures of urban and olive-oil mill wastewaters used as culture medium. Applied Energy. 2013;104:345-52.
- Apandi NM, Mohamed RMSR, Latiffi NAM, Rozlan NFM, Al-Gheethi AAS. Protein and Lipid Content of Microalgae *Scenedesmus sp.* Biomass Grown in Wet Market Wastewater. MATEC Web of Conferences. 2017;103:06011. doi: 10.1051/matecconf/201710306011.
- Makareviciene V, Vaida A, Virginija S, Jurate K. Cultivation of Microalgae Chlorella sp. and Scenedesmus sp. as a Potential Biofuel Feedstock. Environmental Research, Engineering and Management. 2011;57(3):21-7.
- 95. Dawah AM. Efficiency of Inoculating the Green Algae. 2007;11(3):115-25.
- Barrow C, Shahidi F. Marine nutraceuticals and functional foods. CRC Press, United State. 2008.
- Mohammed ZA. Polysaccharides as a protective response against microcystin-induced oxidative stress in *Chlorella vulgaris* and *ScenedeBusmus quadricauda* and their possible significance in the aquatic ecosystem. Ecotoxicol. 2008;17(6):504-16.
- Liu L, Pohnert G, Wei D. Extracellular Metabolites from Industrial Microalgae and their Biotechnological Potential: A Review. Marine Drugs MDPI. 2016;14(10):191. doi.10.3390/md14100191.
- Mendiola JA, Torres CF, Martin A, Santoyo PJ, Tore SA. Use of supercritical CO2 to obtain extracts with antimicrobial activity from *Chaetoceros muelleri* microalgae: A correlation with their lipidic content. Eur Food Res Technol. 2007;224(4):505-10.
- Pratt R, Daniels TC, Eiler JB, Gunnison JB, Kumler WD, et al. Chlorellin, an antibacterial substance from Chlorella. Science. 1944;99:351-2.
- 101. Mayer AMS, Hamann MT. Marine pharmacology in 2001-2002: marine compounds with antihelminthic, antibacterial, anticoagulant, antidiabetic, antifungal, anti-inflammatory, antimalarial, antiplatelet, antiprotozoal, antituberculosis and antiviral activities: affecting the cardiovascular, immune and nervous system and other miscellaneous mechanisms of action. Comparative Biochemistry and Phycology, Part C. 2005;140(3-4): 265-86.
- 102. Dineshkumar R, Narendran R, Jayasingam P, Sampathkumar P. Cultivation and Chemical Composition of Microalgae Chlorella vulgaris and its Antibacterial Activity against Human Pathogens. J Aquac Mar Biol. 2017;5(3):00119. Doi: 10.15406/jamb.2017.05.00119.
- Wilkie AC, Mulbry WW. Recovery of dairy nutrients by benthic freshwater algae. Bioresource Technology. 2002;84(1):81-91.
- Rawar I, Kumar RR, Mutanda T, Bux F. Dual Role of microalgae: Phycoremediation of domestic wastewater and biomass production for sustainable biofuels production. Applied Energy. 2011;88(10):3411-24.
- Ben-Amotz, A. Industrial production of microalgal cell-mass and secondary products- major industrial species *Dunaliella*. In Handbook of Micro Algal Culture Biotechnology and Applied Phycology. Blackwell Publishing Ltd. 2004;273-80.
- Oren A. A Hundred years of *Dunaliella* research: 1905-2005. Saline Systems. 2005;1(1):2.
- Pulz O, Scheibenbogen K, Grob W. Biotechnology with cyanobacteria and microalgae: A Multi-Volume Comprehensive Treatise Biotechnology Germany. Wiley-VCH Verlag GmbH. 2001;10:107-36.
- Lee YK. Microalgal mass culture systems and methods: their limitation and potential. Journal of Applied Phycology. 2001;13(4):307-15.
- Mutsumi T, Karseno TY. Effect of Salt Concentration on Intracellular Accumulation of Lipids and Triglycerides in Marine Microalgae *Dunaliella* cells. Journal of Bioscience and Bioengineering. 2006;101(3):223-6.
- Sun M, Qian K, Sun N, Chang H, Liu J, Shen G. Foot and mouth disease virus VP1 Protein fused with cholera toxin B subunit expressed in *Chlamydomonas reinhardtii* chloroplast. Biotechnology Letters. 2003;25(13):1087-92.
- Geng D, Wang Y, Wang P, Li W, Sun Y. Stable expression of hepatitis B surface antigen gene in *Dunaliella salina* (Chlorophyta). Journal of Applied Phycology. 2003;15(6):451-6.

Cite this article: Geraldine HAS, Baldia SF, Cordero PAJ. Review on the Historical Development of Phytoplankton in the Philippines and their Biological Importance Throughout the Years. Asian J Biol Life Sci. 2020;9(2):239-45.