Research Article

Isolation, Biochemical Characterization of *Rhizobium* sps SNo1 Strain from Root Nodules of *Mimosa pudica* and their Impact on Agriculture Crops

Soundarya Shree KR¹, Nagalambika Prasad¹, Prakruthi G,² Siddalingeshwara KG,² Sharangouda J Patil^{3,*}

¹Department of Microbiology, School of Life Sciences, JSS Academy of Higher Education and Research, Mysuru, Karnataka, INDIA. ²Scientific and Industrial Research Centre, Bengaluru, Karnataka, INDIA.

³Department of Zoology, NMKRV College for Women (Autonomous), Bengaluru, Karnataka, INDIA.

Submission Date: 05-03-2022; Revision Date: 27-03-2022; Accepted Date: 09-04-2022.

ABSTRACT

The present study deals with the isolation of *Rhizobium* bacteria from root nodules of *Mimosa pudica*, and to check its efficiency as bio-fertilizer in improving plant growth. *Rhizobium* sps SN01 was isolated from root nodules of *Mimosa pudica* and confirmed to be *Rhizobium* bacteria through following biochemical tests of indole, methyl red, Voges-Proskauer and citrate utilization (IMViC), urease, hydrogen sulphide, carbohydrate fermentation tests. *Rhizobium* sps SN01 was analysed for antibiotic sensitivity test towards three different antibiotics such as Azithromycin, Amoxicillin and Cephalosporin. To check the efficiency, seeds of legume crops like cow peas, peas, and fenugreek were treated with *Rhizobium* sps, SN01, sowed in soil and coco-peat to study its germination rate and effectiveness in plant growth resulting in root length, shoot length, protein content and chlorophyll content. Impact of *Rhizobium* sps SN01 bacteria in soil and coco-peat quality and improvement was also focused by analyzing micro and macro nutrients responsible for its quality to improve plant growth.

Keywords: *Rhizobium* sps SN01, *Mymosa pudica*, Bio-fertilizer, Legume crops, Cow peas, Soil, Coco-peat.

INTRODUCTION

Bio-fertilizers are actually the natural mini- fertilizers which are responsible for providing safer plant nutrition and increasing the soil fertility improvement through natural processes.^[1]

A bio-fertilizer is a substance containing living microorganisms which was applied to seeds, plant surfaces or soil, colonize the inner part of the plant or the rhizosphere, and also promote growth parameters by increasing the supply or availability of primary nutrients

SCAN QR CODE TO VIEW ONLINE					
	www.ajbls.com				
	DOI: 10.5530/ajbls.2022.11.28				

to the host plant. Bio-fertilizers are cost-effective and ecofriendly in nature. The use of bio-fertilizer improves soil fertility by fixing atmospheric nitrogen, solubilising insoluble phosphates and producing plant growthpromoting substances in the soil. It also promotes nodulation efficiency and increases yield o crops by around 16-60%.^[2] Nitrogen is the essential component which serves as the building blocks of proteins and nucleic acids and it is abundantly found in the earth's atmosphere but it cannot be utilized by plants because of the inert nature due to the presence of triple bonds between the nitrogen atoms. So for the nitrogen to be used by plants it must be fixed or converted to the form of ammonium or nitrite ions. There are certain microorganisms which are capable of converting the atmospheric nitrogen into ammonia or nitrite ions by a process known as nitrogen fixation. These

Correspondence: Dr. Sharangouda J Patil,

Associate Professor, Department of Zoology, NMKRV College for Women (Autonomous), Bengaluru-560041, Karnataka, INDIA.

Email: shajapatil@gmail. com

nitrogen fixation can be done by certain soil microbes symbiotically as well as non-symbiotically.^[1,3]

Many species of leguminous plants form a symbiotic association with various bacteria which are collectively named as "rhizobia". The rhizobia fix inert atmospheric nitrogen (N_2) into biologically useful forms within legume root nodules in a process called "Biological N_2 fixation" (BNF).^[4] This symbiotic association is the largest natural source of the N cycle to sustain natural systems.^[5] Nitrogen fixing bacteria popularly known as *Rhizobium* and it has been found to be having a greatest bio-activity in fixing this atmospheric nitrogen for plants and also increasing the soil fertility.^[6] The aim of the current study is to isolate, characterize the *Rhizobium* bacteria from root nodules of *Mimosa pudica* and to check its impact on agriculture crops as bio-fertilizer in various leguminous plants.

MATERIALS AND METHODS

Isolation of *Rhizobium* bacteria from root nodules of *Mimosa pudica*

Root nodules of *Mimosa pudica* were freshly collected in sample collecting pouches and brought to the laboratory within 24 hr for the isolation of *Rhizobium* bacteria. Root nodules were cleansed thoroughly with tap water, followed by ethanol, distilled water and hydrogen peroxide to remove the adhering contaminants. Series of inoculum obtained from root nodules of *Mimosa pudica* and it was inoculated onto Selective Yeast Extract Mannitol Agar (YEMA) (Containing g/lt of Mannitol-10g; Magnesium Sulphate – 0.2g; Sodium Chloride-0.10g; Potassium di-hydrogen phosphate-0.50g; Calcium chloride-0.20g; ferric chloride-0.01g; Yeast extract-1gm; Congo red – 0.025g: Agar-10gm) and incubated at 35°C for 48 hr.^[5,7]

Biochemical Characterization of Rhizobium sps

Isolates obtained from root nodule of *Mimosa pudica* were subjected to few of the biochemical tests, such as Gram's staining, catalase test, oxidase test, IMViC tests, urease test, hydrogen sulphide test, starch hydrolysis test, carbohydrate fermentation test (Glucose and Lactose) and antibiotic sensitivity test.^[8]

Seed Treatment of Legume Crops with *Rhizobium* sps

The *Rhizobium* sps isolated from *Mimosa pudica* were batch cultured using yeast extract mannitol agar broth (YEMB), after the completion of incubation period, the bacterial load produced was separated by filtration and centrifugation and used as a bio-fertilizer on agriculture legume crops like, fenugreek, peas and cowpea. The above said seeds of legume crops were collected, brought into laboratory, processed by surface sterilization using tap water followed by hydrogen peroxide and distilled water to remove the contaminants adhering to the surface of seeds,. Later seeds were treated with the *Rhizobium* sps and sowed in soil and coco-peat to know the germinate rate. The test was repeated for the other batch of seeds without treating with *Rhizobium* sps and maintained as control.^[9-10]

Evaluation of germination rate, protein and chlorophyll content in the legume crops

The treated and untreated legume crops were evaluated based on germination rate, protein content and chlorophyll content periodically to check the effect of *Rhizobium* sps.

Germination rate of plants in treated batch and untreated batch were studied by measuring its root length and shoot length every 5 days of interval for up to 30 days.^[11]

Protein content was determined as described by Lowry's method^[12] for the duration of 50 days study with interval of 10 days in each batch of crops and the total chlorophyll content was determined by the determination of chlorophyll A and B as described by Patel *et al.*^[13]

Impact of *Rhizobium* sps on nutritional factors present in soil and coco- peat required for plant growth

The nutritional factors like pH, conductivity, organic carbon, organic matter, organic nitrogen, calcium, magnesium, sodium, potassium, nitrate, sulphate, phosphate and micronutrients like iron, zinc, nickel, lead, copper were studied in soil and coco-peat analysed as described by Gomare *et al.*,^[9] with slight modification, in order to check *Rhizobium* sps contribution in enhancement of soil quality and coco-peat quality.^[14]

RESULTS

Isolation of *Rhizobium* Bacteria from Root Nodules of *Mimosa pudica*

Rhizobium sps, was isolated on yeast extract mannitol agar (YEMA), distinct colonies similar to *Rhizobium* sps confirmed by colony characters like circular shape, semi-translucent, raised, whitish in color and designated as SN01 species number. The culture strain was subcultured and stored at 4°C for further work.

Biochemical characterization of Rhizobium sps

The results of biochemical tests are as tabulated in the Table 1 and images of tests are represented in Figures 1 to 5. Biochemical characterizations tests of SN01 showed gram negative rod shaped structure in

Table 1: Results of SN01 strain towards biochemical tests.						
SI. No.	Biochemical tests	Results				
1.	Gram's Staining	Negative				
2.	Shape	Rod				
3.	Catalase test	Positive				
4.	Oxidase test	Positive				
5.	Indole production test	Positive				
6.	Methyl red test	Positive				
7.	Voges Proskauer test	Positive				
8.	Citrate utilization test	Positive				
9.	Hydrogen sulfide production test	Negative				
10.	Carbohydrate fermentation	i test				
a.	Glucose	Positive				
b.	Lactose	Negative				
11.	Urease test	Positive				
12.	Starch hydrolysis test	Positive				

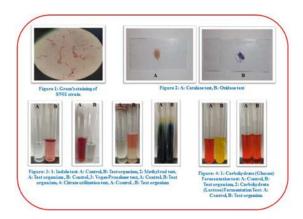




Figure 1: Gram's staining of SN01 stain.

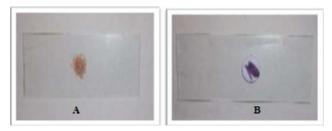


Figure 2: Catalase test, B: Oxidase test

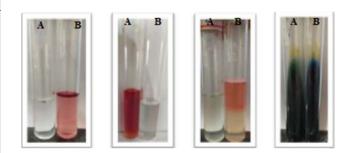


Figure 3: 1 Indole test, A: Control, B: Test organism,
2: Methylred test, A: Test organism, B: Control,
3: Voges-Prouskaer test, A: Control, B: Test organism,
4: Citrate utilization test, A: Control, B: Test organism.

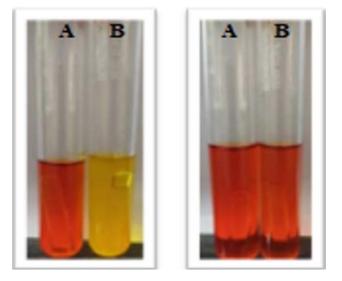


Figure 4: 1: Carbohydrate (Glucose) Fermentation test: A: Control, B: Test organism, 2: Carbohydrate (Lactose) Fermentation test: A: Control, B: Test organism



Figure 5: Antibiotic sensitivity test A: Azithromycin-5mm, B: Cephalosporin-15mm, C: Amoxicillin-Resistant.

Asian Journal of Biological and Life Sciences, Vol 11, Issue 1, Jan-Apr, 2022

Table 2: Effectiveness of <i>Rhizobium</i> sps SN01 in increasing germination rate at 15 th day.									
Potting Mix with Coco-peat									
Legume Crop ↓ [−]	Root Length (cm)		Shoot Length (cm)		Germination Rate (%)		Vigor Index		
	Treated	Untreated	Treated	Untreated	Treated	Untreated	Treated	Untreated	
Cow Pea	15	08	25	18	98	90	2465	1628	
Fenugreek	10	8.5	15	10	96	86	1450	868.5	
Peas	15.3	10	20.2	15	98	88	1994.9	1330	
	Potting Mix with Soil								
Legume	Root Length in cm		Shoot Length in cm		Germination Rate (%)		Vigor Index		
Crop ↓	Treated	Untreated	Treated	Untreated	Treated	Untreated	Treated	Untreated	
Cow Pea	12.4	08	22.6	18.8	98	88	2227.2	1662.4	
Fenugreek	8	07	10	09.2	98	86	988	798.2	
Peas	14	10.1	18.5	12.4	98	88	1827	1101.3	

Table 3: Effectiveness of <i>Rhizobium</i> sps SN01 in increasing protein and chlorophyll content at 15 th day.								
Potting Mix	Coco-peat			Soil				
→ Legume Crop ↓	Protein Content (mg/g) Chlorophyll Content (mg/g)		Protein Content (mg/g)		Chlorophyll Content (mg/g)			
	Treated	Untreated	Treated	Untreated	Treated	Untreated	Treated	Untreated
Cow Pea	59.5	52.0	0.64	0.48	40.0	39.5	0.43	0.6
Fenugreek	54.0	42.0	0.4	0.3	34.0	13.5	0.29	0.21
Peas	39.0	30.5	0.51	0.41	49.5	38.0	0.5	0.46

Gram's staining studies, positive result was showed by strain SN01 for catalase test, oxidase test, IMViC tests, urease test and starch hydrolysis test, in carbohydrate fermentation test strain SN01 showed positive result for glucose and negative result for lactose. Negative result was recorded for triple sugar iron agar test by SN01 strain. Antibiotic sensitivity test for antibiotics amoxicillin, cephalosporin, azithromycin was tested, strain SN01 showed 05mm and 15mm of inhibition towards azithromycin and cephalosporin respectively, whereas strain SN01 was resistant for amoxicillin.

Seed treatment of legume crops with *Rhizobium* sps SN01

Healthy seeds of cow peas, fenugreek, peas were selected for the germination study, the seeds were treated with culture of *Rhizobium* sps SN01 and sowed in two set of potting trays, and the other set was control which was untreated with *Rhizobium* bacteria, two sterile potting mixes, soil and coco-peat were used in the study.

Evaluation of Germination Rate, protein and chlorophyll Content in the Legume Crops

Rhizobium sps SN01 effectiveness in improving plant germination rate and plants major character like protein and chlorophyll content were studied for a period of 30 days. Germination rate and its vigor index was studied by checking root and shoot length of treated and untreated legume crops, Shoot and root length was more in treated plants compared to untreated plants, out of peas, cowpea and fenugreek plants, cowpeas in both soil and coco-peat showed higher vigor index. The results are as tabulated in Table 2, 3 and image in Figure 6. The Protein and chlorophyll content was measured in the plants, it was found to be treated plants have more protein and chlorophyll content compared untreated legume crops in both potting mix.

Impact of *Rhizobium* sps on nutritional factors present in soil and coco-peat required for plant growth

Nutritional factors in potting mixes play significant role in maintaining its quality and it also improves plant growth, So *Rhizobium* sps SN01 activity towards increasing nutritional factors were studied, major and minor nutrients in soil and coco-peat before treatment and after treatment were analysed, from the study it was noticed that *Rhizobium* sps SN01 was responsible in increasing the required nutritional factor in both soil and coco-peat. The results of the study are represented in Table 4.



Figure 6: Germination rate in legume crops, A: Cow pea, B: Fenugreek, C: Peas.

nutritional elements.							
	S	oil	Coco	-peat			
Nutritional Elements	Treated	Untreated	Treated	Untreated			
PH	6.8	6.5	5.5	5.2			
Conductivity (µs/cm)	204	198	494	406			
Organic Carbon (%)	1.0	0.8	8.7	08			
Organic Matter (%)	1.7	1.0	15	11			
Organic Nitrogen (mg/kg)	0.28	0.2	30.1	25.2			
Calcium (mg/kg)	20	16	50	36			
Magnesium (mg/kg)	10	08	10	08			
Total Nitrogen (mg/ kg)	4.8	1.2	39	30.8			
Sulphate (mg/kg)	408	398	188	115			
Sodium (mg/kg)	22	18	60	48			
Potassium (mg/kg)	90	82	120	86			
Phosphate (mg/kg)	10.2	08	16	12			
Lead (mg/kg)	31	25	10	4			
Copper (mg/kg)	19	10	11	6			
Iron (mg/kg)	1748	1563	1555	1089			
Nickel (mg/kg)	15	10	05	0.9			
Zinc (mg/kg)	38	28	33	22			

Table 4: Impact of Rhizobium sps SN01 in increasing

DISCUSSION

Enrichment of soil nutrients by nitrogen fixing bacteria in leguminous plants as a symbiotic relationship has been known history and natural system. Scientific investigation on this symbiotic relationship of plants and microbes was reported from 19th century and it evidenced the presence of microbial diversity in legume root nodules in that bacteria's were predominant and responsible for fixing atmospheric nitrogen.^[3] Rhizobium spp. considered well known bacteria for the symbiotic relationshion with various plants to fix the nitrogen as a

primary source. Such types of bacteria are infecting the leguminous plant roots, leading to establish the lump formation or cystic nodules where the occurrence of synthesis of nitrogen fixation.

Isolation and characterization of Rhizobium strain assessed for various studies, in that, the biochemical test conducted strain SN01 was confirmed to be a species of Rhizobium by the characters like, shape, size, media, pH, temperature, substrate specificity, etc. Hence the strain was further designated as Rhizobium sps SN01 with the result remarks. The results obtained for the test organism was similar to the work of Kumar et al.[15] and Purwaningsih et al.[16]

The work of Patel et al.^[13] on soyabean with Rhizobium sps and other bio-fertilizers in accordance with germination rate, vigor index, protein and chlorophyll content proved that Rhizobium is an effective bio-fertilizer in improving and maintaining plant growth. Ranjbar et al.[11] proved the effectiveness of Rhizobium leguminosarum on Pisum satvium in increasing seed yield rate and various physical properties. Hence these similar reports are evidence for our work to present that Rhizobium sps SN01 strain plays a vital role in improving plant growth.

Nutrition in potting mixes play significant role in maintaining its quality and it also improves plant growth, So Rhizobium sps SN01 activity towards increasing nutritional factors were studied, major and minor nutrients in soil and coco-peat before treatment and after treatment were increased all the parameters such as pH (6.8 & 5.5), conductivity (204 & 494 μ s/cm), organic carbon (1.0 & 8.7%), organic matter (1.7 & 15%), organic nitrogen (0.28 & 30.1 mg/kg), calcium (20 & 50 mg/kg), magnesium (10 & 10 mg/kg), total nitrogen (4.8 & 39 mg/kg), sulphate (408 & 188 mg/kg), sodium (22 & 60 mg/kg), potassium (90 & 120 mg/kg), phosphate (10.2 & 16), lead (31 & 10 mg/kg), copper (19 & 11 mg/kg), iron (1748 & 1555 mg/kg), nickel (15 & 5 mg/kg), zinc (38 & 33 mg/kg) compared tocontrol untreated parameters respectively. Gomare et al.^[9] and Mia et al.^[10] reported on rice and legume seeds respectively to check the ability of Rhizobium sps to help plant growth.

The present work also shows similar results and proves that Rhizobium sps SN01 is effectively improving germination rate of legume crops like cow peas, peas and fenugreek in soil and coco-peat conditions. Similar observation of Gomare et al.^[9] proven that Rhizobium sps isolated from root nodules are responsible in enhancing soil characteristics before adding inoculum and after adding inoculum.

CONCLUSION

In the present study Rhizobium sps SN01 isolated from root nodules of Mimosa pudica was efficient in improving germination rate of legume crops like cow peas, peas and fenugreek in soil and coco-peat conditions, out of three crops Rhizobium sps SN01 was more responsible towards cowpeas in improving germination rate to 98% in soil and coco-peat, vigor index to 2465 in coco-peat and 2227.2 in soil, protein to 595mg/g in coco-peat and 400 mg/g in soil, chlorophyll to 0.64 mg/g in coco-peat and 0.43 mg/g in soil. Rhizobium sps SN01 influence in improving soil quality and coco-peat quality was also more effective, it was found that all the micro and macro nutrients required for plant growth was found to be significant in higher concentration compared to untreated soil and coco-peat. The present study concludes that potting mix coco-peat is more suitable for the growth of legume crops in the presence of Rhizobium sps SN01, in acquiring more yield and quality of all the three legume crops cow peas, peas and fenugreek.

ACKNOWLEDGEMENT

Department of Microbiology, JSS Academy of Higher Education and Research, Mysuru, Karnataka, India, provided basic facilities to the authors. We thank to Dr. Siddalingeshwara KG, R&D, Head, Scientific and Industrial Research Centre, Bengaluru, Karnataka, India for providing plant growth work facility and assessment.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- Roychowdhury D, Paul M, Banerjee SK. Isolation identification and characterization of bacteria (Rhizobium) from chickpea (*Cicer arietinum*) and production of biofertilizer. Eur J Biotechnol Biosci. 2015;3:26-9.
- Htwe AZ, Moh SM, Moe K, Yamakawa T. Biofertiliser Production for Agronomic Application and Evaluation of Its Symbiotic Effectiveness in Soybeans. Agronomy;9(4). doi: 10.3390/agronomy9040162.
- Zahran HH. Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. Microbiol Mol Biol Rev. 1999;63(4):968-89. doi: 10.1128/MMBR.63.4.968-989.1999, PMID 10585971.
- Peoples MB, Herridge DF, Ladha JK. Biological nitrogen fixation: An efficient source of nitrogen for sustainable agricultural production? Plant Soil. 1995;174(1-2):3-28. doi: 10.1007/BF00032239.
- Datta A, Singh RK, Tabassum S. Isolation, characterization and growth of Rhizobium strains under optimum conditions for effective biofertilizer production. Int J Pharm Sci Rev Res. 2015;32(1):199-208.
- Randa HE, Mohamed SS, Sherif AM, Osman AG. Rhizobium biofertilizer (Okadin) production and future prospects in Sudan, Environ Nat Res. Int J. 2016;1(1):1-12.
- 7. Wagner SC. Biological nitrogen fixation. Nat Educ Knowl. 2011;3(10):15.
- Aneja KR. Experiments in microbiology, plant pathology, tissue culture and microbial biotechnology, new age international publishers. 4th ed; 2003.
- Gomare KS, Mese M, Shetkar Y. Isolation of Rhizobium and cost-effective production of biofertilizer. Ind J Life Sci. 2013;2(2):49-53.
- Mia MAB, Shamsuddin ZH. Rhizobium as a crop enhancer and biofertilizer for increased cereal production. Afr J Biotechnol. 2010;I;9(37):6001-9.
- Ranjbhar SN, Kaul HP, Desalegn G, Wienkoop S. Rhizobium impacts on seed productivity, quality, and protection of Pisum satvum upon disease stress caused by Didymella pinodes: Phenotypic, proteomic and metabolic traits, Front Plant. Sci J. 2017;8. doi: doi=10.3389/fpls.2017.01961.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. J Biol Chem. 1951;193(1):265-75. doi: 10.1016/ S0021-9258(19)52451-6. PMID 14907713.
- Patel S, Bafna A, Maheshwari RS, Rangwala T. Comparative study of impact of Rhizobium, phosphate solubilizing bacteria, vermicompost and urea fertilizer on growth parameters, chlorophyll ad protein content of soyabean (Glycine max). Int J Res Biosci. 2016;5(1):43-5.
- 14. Soil Analysis Manual, Rokupur agricultural research centre (RARC) and Japan international co-operation agency (JICA); 2014.
- Kumari NB, Nagaraju B, Mallikarjuna K. Biochemical characterization and protein profile by SDS- PAGE of French bean (*Phaseolus vulgaris* L.) associated Rhizobia, Innovat. Int J Med Pharm Sci. 2017;2(2):8-13.
- Purwaningsih S, Agustiyani D, Antonius S. Diversity, activity, and effectiveness of *Rhizobium* bacteria as plant growth promoting rhizobacteria (PGPR) isolated from Dieng, central Java. Iran J Microbiol. 2021;13(1):130-6. doi: 10.18502/ijm.v13i1.5504, PMID 33889372.

Cite this article: Soundarya SKR, Prasad N, Prakruthi G, Siddalingeshwara KG, Patil SJ. Isolation, Biochemical Characterization of *Rhizobium* sps SN01 Strain from Root Nodules of *Mimosa pudica* and their Impact on Agriculture Crops. Asian J Biol Life Sci. 2022;11(1):200-5.